

Multiresolution Techniques for the Classification of Bioimage and Biometric Datasets

Amina Chebira¹ and Jelena Kovačević^{1,2}

¹ Center for Bioimage Informatics and Dept. of Biomedical Engineering,

² Dept. of Electrical and Computer Engineering,
Carnegie Mellon University, Pittsburgh, PA, USA

ABSTRACT

We survey our work on adaptive multiresolution (MR) approaches to the classification of biological and fingerprint images. The system adds MR decomposition in front of a generic classifier consisting of feature computation and classification in each MR subspace, yielding local decisions, which are then combined into a global decision using a weighting algorithm. The system is tested on four different datasets, subcellular protein location images, drosophila embryo images, histological images and fingerprint images. Given the very high accuracies obtained for all four datasets, we demonstrate that the space-frequency localized information in the multiresolution subspaces adds significantly to the discriminative power of the system. Moreover, we show that a vastly reduced set of features is sufficient. Finally, we prove that frames are the class of MR techniques that performs the best in this context. This leads us to consider the construction of a new family of frames for classification, which we term lapped tight frame transforms.

1. BACKGROUND AND MOTIVATION

In this work, we show that, by looking into four different biological/biometric applications, the underlying problem is classification and thus, an accurate and efficient algorithm would be of great use. We proceed to give a general background on classification and how it was used in the first problem, that of recognizing proteins based on the images depicting their location within the cell. As in that attempt, the authors had great success with the simplest of MR techniques, we postulate that using more sophisticated ones would lead to more accurate classification. We thus proceed to review nonredundant MR tools—MR bases, and show how, in their adaptive incarnation, they have been used with great success in fingerprint recognition. In the same problem, the authors observed that the translation variance of these bases might pose a problem and suggest considering redundant MR techniques—frames, which we then review.

1.1. Motivation: Classification Problems in Bioimaging and Biometrics

Systems biology entails the study of the interactions between the components of a biological system and the mechanisms by which these interactions give rise to the function and behavior of that system. It can be viewed as a “macro” approach that encompasses mathematical and computational modeling based on quantitative data collected within each component of the biological system.

The Need for Automated Processing. Advances in biochemistry, probes, and microscopy gave the biologists the opportunity to observe cells and cell processes at a level never seen before, which led to the collection of huge amounts of 2D, 3D and even higher-dimensional data. As a result, visual inspection of these datasets, always error-prone, nonreproducible and subjective, became impractical as well. Hence the need for automated, accurate and efficient systems to extract knowledge contained in the collected data.

Classification of Biological and Biometrics Images. Such automated knowledge extraction requires the expertise developed in signal processing, machine learning and mathematics. In the project of determination of protein subcellular location patterns, Murphy et al. identified classification as the underlying problem [1]. Similarly, in the project of determination of developmental stages in fly embryos [2], we realized that the problem is again that of classification. Not surprisingly then, in several other projects, such as the development of teratomas in stem cells, where multiple tissues are present and need to be recognized, as well as fingerprint recognition, the need for classification emerged. Thus, an accurate and efficient algorithm for classification would be of great use to biologists, motivating the developments in this work.

Determination of Protein Subcellular Location Patterns. The field of proteomics entails the study of proteins and their role and function in various cellular mechanisms. One of the critical characteristics of a protein is its subcellular location (PSL), that is, its spatial distribution within the cell. Knowledge of the location of all proteins will be essential for building accurate models that capture and simulate cell behavior, and eventually can be expected to be useful for early diagnosis of disease and/or monitoring of therapeutic effectiveness. Once successfully tagged (using green fluorescent protein (GFP) for example), the cells are imaged using one of many models of fluorescence microscopes to produce a multidimensional dataset. Murphy et al. pioneered the use of automated systems for protein identification based on their subcellular location patterns [1].

The Need for Classification. Given that mammalian cells are believed to express tens of thousands of proteins, comprehensive analysis of protein location requires acquisition of images whose numbers are beyond our ability to analyze visually. Moreover, as today's microscopes allow for imaging of high-dimensional datasets, both the enormous volume as well as the high dimensionality of the data render human analysis time-consuming, prone to error and ultimately, impractical, leading to the "holy grail" for PSL bioimage interpretation and analysis: develop a system for fast, automatic, and accurate recognition of proteins based on their subcellular location.

Detection of Developmental Stages in *Drosophila* Embryos. *Drosophila*, also known as small fruit fly, is an important model organism in developmental biology, as it shares cellular and molecular processes with higher eukaryotes including humans. To fully comprehend such mechanisms, scientists study genome sequences provided by the genome projects. The genome sequence of *Drosophila melanogaster* was published in 2000 [3]. In the project led by Minden at CMU, the formation of the ventral furrow is observed in early embryonic development as it is known to be a key morphogenic event. Once a gene is silenced using RNAi, the fly embryos are tagged with GFP and imaged using a confocal laser fluorescence microscope. This acquisition process allows for the collection of z-stack images of embryo volumes in time [2].

The Need for Classification. *Drosophila* gastrulation involves four major morphogenic events, the first one being ventral furrow formation, which can be divided into three steps: (1) The initial stage when no ventral furrow is formed yet. (2) During Stage 2, the entire ventral furrow collapses inward, bringing a band into the interior of the embryo over a period of several minutes. (3) Stage 3 consists of having a closed and formed ventral furrow. To study this process, Minden et al. acquire 3D volumes of the formation of the ventral furrow in time. As manual processing of huge amounts of these 4D (space + time) datasets is impractical, cumbersome and error-prone, we believe that reliable, accurate, and efficient algorithms for automated 4D image and phenotype analysis are crucial to enable high-throughput functional genomics screens such as this one.

Classification of Histological Stem-Cell Teratomas. The study of stem cells is one of the most exciting and promising research areas in the biomedical field. Stem cells are unspecialized cells that have the remarkable ability to divide, renew for long periods of time and become specialized (differentiate). The mechanisms that lead such cells to divide, remain undifferentiated or specialize are yet to be understood.

In the project led by Castro and Ozolek at the University of Pittsburgh Medical School, the aim is to answer some of these questions using high-field magnetic resonance imaging (MRI) and histological staining methods. First, stem cells are implanted in the testis of a rat. Then, with no control over the differentiation process, the implanted stem cells develop into various tissue types such as muscle, epithelial and bone. This tissue-rich tumor is then removed and imaged using MRI as well as stained with Hematoxylin and Eosin (H&E) and then imaged. An important step towards understanding stem-cell division and differentiation would be to coregister cellular class identifications from the highly-detailed histological images to the difficult-to-read MRI data. The goal here is to enhance the diagnostic capabilities of MRI using histology images as ground truth.

The Need for Classification. Currently, the 2D MRI and histological images are acquired and manually interpreted by experts. That is, each image is segmented into meaningful regions such as bones and muscle, and those regions are appropriately labeled. A large amount of data begs for an automated and accurate system that will perform these tasks efficiently. The process of labeling mentioned above is in fact a classification problem again.

Fingerprint Recognition. Personal identification has been a topic of interest for some time, with various solutions proposed. Accessing buildings or facilities, withdrawing money or using a credit card, gaining access to electronic information on a local computer or over the Internet, are all examples of situations which require accurate and reliable methods of personal identification, and solutions vary greatly. Using human biometric

characteristics (fingerprints, irises, faces, etc) has great advantages over other modalities: the information cannot be lost or forgotten, and forgery requires greater skill.

The Need for Classification. The most familiar and studied modality of biometric recognition is the fingerprint. Because acquisition of fingerprint images is minimally invasive and requires little hardware (ink, paper and a digital camera are the minimum requirements), fingerprint recognition is a highly researched field. By recognition here we mean identification of a person (namely one individual corresponds to one class)—again a classification problem. A crucial goal in processing this biometric data is to do so automatically, accurately and fast.

1.2. Classification

As we have seen above, the need for classification of biological datasets is pervasive, and ever better solutions are needed. Thus, we start with a brief overview of classification methods.

Problem Statement. The problem we are addressing is that of classifying images from biological and biometric datasets. Assume that the images are of size $N \times N$ and let \mathbb{R} denote the set of intensities covered by all the images in the given dataset. Thus, the classification problem can be formulated as designing a map from the *signal space* of the examined images $\mathcal{X} \subset \mathbb{R}^{N \times N}$, to a *response space* $\mathcal{Y} \subseteq \{1, 2, \dots, C\}$ of class labels. The decision d is the map, $d: \mathcal{X} \mapsto \mathcal{Y}$ that associates an input image with a class label. To reduce the dimensionality of the problem, one sets up a feature space $\mathcal{F} \subset \mathbb{R}^f$, $f \leq N^2$, between the input space and the response space. The feature extractor θ is the map $\theta: \mathcal{X} \mapsto \mathcal{F}$, and the classifier ν is the map $\nu: \mathcal{F} \mapsto \mathcal{Y}$. The goal is to find a (θ, ν) pair that maximizes the classification accuracy.

Feature Extraction. Features are numerical descriptors that characterize the input data, usually in a lower-dimensional space. We focus on the following feature sets:

Haralick Texture Features (T_1 with 13 features or T_2 with 26 features), are calculated using four co-occurrence matrices which are combined in various ways to give either 13 or 26 measures [4].

Morphological Features (M , 16 features), visually describe distinctive aspects of images as discerned by the human eye.

Zernike Moment Features (Z , 49 features), computed for an image are similarity measures between the corresponding Zernike polynomials and the image.

Classifiers. We focus here on a specific class of non linear classifiers: Neural networks (NNs), which are classifiers based on grouping the input vectors (features) into intersections of clusters of one type while the union of all such intersections yields the entire feature space.

Determination of Protein Subcellular Location Patterns: Use of Simple MR Techniques. The heart of systems that can classify PSL patterns is a set of features (T_1, T_2, M , and Z) describing the spatial distribution of proteins in each cell image. Of particular relevance to the work described here is the use of simplest MR features such as wavelet (30 features) and Gabor (60 features) features, as the addition of these resulted in a significant improvement of classification accuracy to 91.5% for the 2D HeLa dataset [1].

The Need for Multiresolution. As the introduction of the simplest MR features produced a statistically significant jump in classification accuracy, our hypothesis is that more sophisticated MR techniques would result in even more accurate classification. In particular, the three crucial characteristics of MR [5] we wish to explore are: (a) localization, (b) adaptivity and (c) fast and efficient computation. These are some of the major reasons for the phenomenal success of MR techniques in real applications and one of the reasons to incorporate MR features into our system.

1.3. Multiresolution Techniques

We now give a brief overview of MR techniques, which have been extensively studied and used in signal and image processing over the past two decades [5]. MR processing means analysis and processing of data at different resolutions and/or scales. MR transforms decompose a signal into zooming spaces (coarse spaces and many detail spaces called *subbands*) and are implemented by *filter banks* (*FBs*), through filtering and sampling.

Nonredundant Multiresolution Techniques: Bases. Most of MR techniques in use are nonredundant—the underlying mathematical structures are *bases* (*MRBs*). Assume finite-dimensional spaces \mathbb{R}^m or \mathbb{C}^m . Given

a basis for such a space, $\Psi = \{\psi_i\}_{i=0}^{m-1}$, we associate to it a matrix (operator) which we will also call Ψ , and which has basis vectors as its columns ($\psi_{i,j}$ is the j th element of the i th basis vector). Given a pair of biorthogonal bases $(\Psi, \tilde{\Psi})$ dual to each other, a signal x belonging to \mathbb{R}^m or \mathbb{C}^m can be expressed as:

$$x = \Psi X = \Psi \tilde{\Psi}^* x, \quad (1)$$

where X is the vector from \mathbb{R}^m or \mathbb{C}^m of so-called *transform* coefficients (inner products), and $\tilde{\Psi}^*$ denotes the Hermitian transpose of the dual basis $\tilde{\Psi}$. If the expansion is into an orthonormal basis (ONB), then $\Psi = \tilde{\Psi}$ and the above becomes $\Psi \Psi^* = I$, which further implies that Ψ is a unitary matrix.

Filter-Bank View of Bases. The only infinite-dimensional class of MR decompositions we discuss here are those implemented by FBs, as these are bases used in applications and our only link to the real world. The vectors (signals) live in the infinite-dimensional Hilbert space $\ell^2(\mathbb{Z})^*$. A FB is the basic signal processing structure used to implement most MR transforms (Fig. 2 depicts a FB with m channels and sampling by n). When $m = n$, we deal with critically-sampled FBs implementing bases. A thorough analysis of FB bases is given in [6].

A FB decomposition can be expressed as in (1) where x is now an infinite sequence belonging to $\ell^2(\mathbb{Z})$, X is an infinite sequence of transform coefficients (inner products) in $\ell^1(\mathbb{Z})$, and Ψ is the basis expansion matrix given in a setting with finite impulse response (FIR) filters. The matrix Ψ is used in the synthesis FB (the reconstruction step) whereas its dual, $\tilde{\Psi}$, is used in the analysis FB (the decomposition step). Assume that the nonzero support of the filter ψ_i , or, its length is $l = km$ (if not, we can always pad with zeros), and write the basis operator as

$$\Psi = \begin{pmatrix} \ddots & \vdots & \vdots & \vdots & \vdots & \vdots & \ddots \\ \cdots & \Psi_0 & 0 & \cdots & 0 & 0 & \cdots \\ \cdots & \Psi_1 & \Psi_0 & \cdots & 0 & 0 & \cdots \\ \cdots & \vdots & \vdots & \vdots & \vdots & \vdots & \cdots \\ \cdots & \Psi_{k-1} & \Psi_{k-2} & \cdots & \Psi_0 & 0 & \cdots \\ \cdots & 0 & \Psi_{k-1} & \cdots & \Psi_1 & \Psi_0 & \cdots \\ \ddots & \vdots & \vdots & \vdots & \vdots & \vdots & \ddots \end{pmatrix}, \text{ with } \Psi_r = \begin{pmatrix} \psi_{0,rm} & \cdots & \psi_{m-1,rm} \\ \vdots & \ddots & \vdots \\ \psi_{0,rm+m-1} & \cdots & \psi_{m-1,rm+m-1} \end{pmatrix}, \quad (2)$$

where $r = 0, \dots, k-1$ and each block Ψ_r is of size $m \times m$. We can rephrase the basis decomposition in the z -domain as well using polyphase analysis. A polyphase matrix $\Psi_p(z)$ collects the subsequences modulo n . For bases, $\Psi_p(z)$ is of size $m \times m$ and can be written as:

$$\Psi_p(z) = \sum_{r=0}^{k-1} \Psi_r z^{-r}, \quad (3)$$

with Ψ_r as in (2). A paraunitary polyphase matrix (representing an ONB) satisfies $\Psi_p(z) \Psi_p^*(z) = cI$, where c is a constant.

Block Transforms. When the filter length l is equal to the sampling factor m , we have a *block transform*. Then, in (2), only Ψ_0 is nonzero, making Ψ block-diagonal. In effect, since there is no overlap between processed blocks, this can be analyzed as a finite-dimensional case, where both the input and the output are m -dimensional vectors. This shows how finite-dimensional bases can be analyzed in the FB context.

Lapped Orthogonal Transforms. In practice, the use of block transforms can produce artifacts known as “blocking effects” (since there is no overlap between the basis functions—processed blocks), and thus solutions were sought with longer basis functions. One such solution is the *Lapped Orthogonal Transform (LOT)*. The LOTs can be seen as a class of m -channel FBs implementing bases, originally developed for filters of length $l = 2m$ and later generalized to arbitrary integer multiples of m [7]. Compared to block transforms, the LOT

*In fact, we can investigate finite-dimensional MR decompositions within the FB framework as well.

keeps the same number of filters but doubles their length, which means that the basis functions of adjacent blocks overlap by half their size, thus removing the blocking effects. However, LOTs are not solely determined by their length, but by the specific form of their basis vectors as well.

In general, for a FB with filter length $l = 2m$, the time-domain matrix Ψ has a double diagonal, that is, in (2), only Ψ_0 and Ψ_1 exist. Thus, (3) reduces to

$$\Psi_p(z) = \Psi_0 + z^{-1}\Psi_1, \quad (4)$$

where $\Psi_r, r = 0, 1$, are $m \times m$ matrices with $(\Psi_r)_{j,i} = \psi_{i,j}$ for $i = 0, \dots, m-1$ and $j = mr, \dots, mr+m-1$. Since the LOT is a unitary transform, that is, $\Psi\Psi^* = \Psi^*\Psi = I$ the following must be satisfied:

$$\Psi_0\Psi_0^* + \Psi_1\Psi_1^* = \Psi_0^*\Psi_0 + \Psi_1^*\Psi_1 = I, \quad (5)$$

$$\Psi_0^*\Psi_1 = \Psi_1^*\Psi_0 = 0, \quad \Psi_0\Psi_1^* = \Psi_1\Psi_0^* = 0. \quad (6)$$

Two main classes of LOTs exist distinguished by whether they use cosines or complex exponentials in their basis functions. We concentrate here on a particular family that uses cosine basis functions:

The Princen-Johnson-Bradley LOT (PJB-LOT). [8] The PJB-LOT basis functions are given by:

$$\psi_{i,j} = \sqrt{\frac{1}{m}} \cos\left(\frac{\pi(2i+1)(2j-m+1)}{4m}\right), \quad i = 0, \dots, m-1 \text{ and } j = 0, \dots, 2m-1. \quad (7)$$

With J and anti-diagonal matrix, and thanks to the particular structure of the cosines:

$$\Psi_0\Psi_0^* = \frac{1}{2}(I - J), \quad \Psi_1\Psi_1^* = \frac{1}{2}(I + J). \quad (8)$$

With this construction, we will have, similarly to the Discrete Fourier Transform (DFT), fixed basis functions allowing no freedom in design. To allow for better designs, one can add a window that multiplies each filter resulting in a modulated FB over the frequency band. This modulated FB can be modeled as $W\Psi$, where the window $W = \text{diag}\{w_j\}_{j=0}^{2m-1}$ is symmetric $w_j = w_{2m-1-j}, j = 0, \dots, 2m-1$. Now, (5) becomes

$$W\Psi_0\Psi_0^*W + JWJ\Psi_1\Psi_1^*JWJ = I. \quad (9)$$

Substituting (8) into (9), we obtain

$$\frac{1}{2}(W^2 + JW^2J) = I, \quad (10)$$

implying that the window has to satisfy $w_j^2 + w_{m-1-j}^2 = 2$, for $j = 0, \dots, m-1$.

Discrete Wavelet Transform and Wavelet Packets. The *Discrete Wavelet Transform (DWT)*, a famous MR tool, is a basis expansion and as such nonredundant (critically sampled). The *dyadic* DWT is built by iterating a two-channel FB with sampling factor $n = m = 2$ on the lowpass channel. One can also build trees by, at each level, iterating on any subset of the branches of the FB. This is known as *Wavelet Packets (WP)*. Both DWTs and WPs can be block transform or not depending on the length of the filters.

1.3.1. Fingerprint Recognition: Use of Nonredundant MR Bases

Due to the nature of fingerprint images, Hennings et al. [9] used an adaptive wavelet packet (WP) approach in combination with correlation filters to solve the recognition problem. The idea was to design a scheme that could adapt itself to the class at hand across multiple scales and resolutions where localized features might be found, clearly calling for the use of WPs. Thus, instead of designing a single correlation filter for a pattern class, a correlation filter was designed for each leaf in the best WP tree found for that class. Finally, if the image belongs to the pattern class of the filter, the correlation plane output contains a sharp peak; if not, no such peak exists. A measure of performance that measures the peak-to-correlation energy (PCE), called *match score*, was designed to discriminate between true and impostor classes. A significant improvement in all classes was achieved by using WPs (98.32% accuracy) compared to the use of correlation filters (81.59% accuracy).

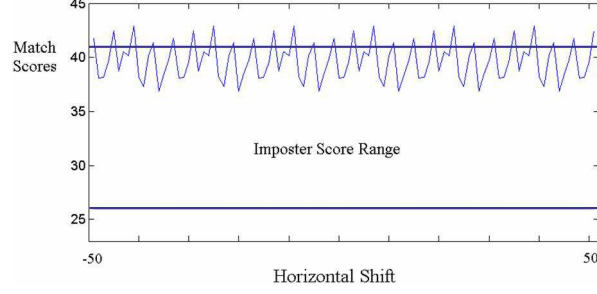


Figure 1. Periodic translation invariance of match scores in a fingerprint recognition system (from [9]).

The Need for Redundant Multiresolution Techniques. Although correlation filters are translation invariant in the image intensity domain, they are not translation invariant in the wavelet domain, as the WPs involve downsampling. The authors in [9] translated images from a difficult and an easy class and computed a match score for each translation. For the easy class, there was still complete separation between authentic scores and the range of impostor scores, despite the translations. When this separation is not as wide, the impostor scores overlap with the match scores, thereby reducing the accuracy of the system (see Fig. 1). This clearly calls for the use of translation-invariant transforms: Redundant MR transforms.

1.3.2. Redundant Multiresolution Techniques: Frames

We start with a brief account of redundant MR techniques which are called *frames* (*MRFs*), first in finite dimensions and then follow up with how signals are represented using frames in infinite dimensions via FBs. An introductory account on frames was written by us [6, 10].

In a finite-dimensional space (\mathbb{R}^n or \mathbb{C}^n), a frame is defined as a set Φ of m frame vectors $\Phi = \{\varphi_0, \dots, \varphi_{m-1}\}$ where m is larger than n . As for bases, we associate to the frame a rectangular matrix of size $n \times m$, also called Φ , that has the frame vectors as its columns.

Similarly to bases, one can check that frames expand signals in \mathbb{R}^n with $x = \Phi X = \Phi \tilde{\Phi}^* x$, where $\tilde{\Phi}$ represents the dual frame. If $\Phi = \tilde{\Phi}$, then we have what is called a *tight frame* (*TF*), and the expansion becomes $\Phi \Phi^* = I$. If all frame vectors have the same norm, the frame is termed an *equal-norm frame* (*ENF*), while if all the norms are equal to 1, this is a *unit-norm frame* (*UNF*). By combining this with the requirement of tightness, we can have *equal-norm tight frames* (*ENTFs*), as well as *unit-norm tight frames* (*UNTFs*).

Filter-Bank View of Frames. In an m -channel FB with sampling factor n , if $m > n$, we deal with an oversampled FB implementing a frame (see Fig. 2).

For a TF, $\tilde{\varphi}_i = \varphi_i$. The FB frame decomposition can be expressed as in (1) (substituting Φ for Ψ), where x is an infinite sequence belonging to $\ell^2(\mathbb{Z})$, X is an infinite sequence of transform coefficients (inner products), and Φ is the frame expansion matrix.

Assuming again that the nonzero support of the filters (frame vectors) length is $l = kn$, we can write the frame operator Φ as in (2), with matrices $\Phi_r, r = 0, \dots, k-1$, being rectangular of size $n \times m$.

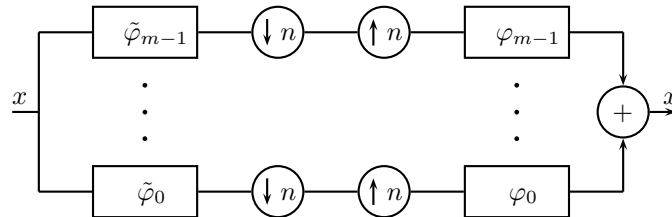


Figure 2. An FB implementation of a frame expansion: It is an m -channel FB with sampling by n .

We can rephrase the frame decomposition in the z -domain as well, where a FB implements a TF decomposition in $\ell^2(\mathbb{Z})$ if and only if its polyphase matrix $\Phi_p(z)$ is paraunitary [11]. For frames, the polyphase matrix $\Phi_p(z)$ is of size $n \times m$ and can be written as in (3) (substituting Φ for Ψ), where Φ_r are of size $n \times m$ as in (2).

Seeding. In an ever-continuing search for new frame families, an appealing option is the process of obtaining TFs from ONBs in larger dimensions, known as the Naimark Theorem or *seeding* [12].

DEFINITION 1.1. We say that a frame Φ is obtained by seeding from a basis Ψ by deleting a suitable set of columns of Ψ . We write $\Phi^* = \Psi[\mathbb{J}]$ where \mathbb{J} is the index set of the retained columns.

All tight frames can be obtained this way. One of the most famous frame families, the Harmonic Tight Frames (HTFs) is obtained by seeding the DFT. In FB parlance, seeding is done on the polyphase matrix. Given $\Psi_p(z)$, the $m \times m$ polyphase matrix associated with a basis of size m , then $\Psi_p(z) = \Psi_0$, and

$$\Phi_p^*(z) = \Phi_0^* = \Psi_p[\mathbb{J}] \quad (11)$$

is the transpose of the frame polyphase matrix.

Block Transforms. When $l = n$, that is, the length of the frame vectors equals the sampling factor, we obtain a block transform. One example of block transform with frames is HTFs.

2. MULTIREOLUTION CLASSIFICATION ALGORITHM

In the last section, we saw that the classification problem is ubiquitous in biology and biometrics, and that MR techniques might make classification more accurate. The results obtained in [9] seem to indicate that adaptive MR techniques, frames in particular, might be needed. Having motivated the use of adaptive MR in classification as well as the need for redundant MR transforms, we now test that hypothesis.

2.1. Multiresolution Classification

We now describe the adaptive MR classification algorithm developed; details and results are described in [13–16].

Main Idea. As argued in Section 1.2, we would like to extract discriminative features within space-frequency localized subspaces. These are obtained by MR decomposition; that is, instead of adding MR features as in [1], we compute features in the MR-decomposed subspaces.

Our initial idea was to use WPs since they adapt themselves to the signal at hand, and just as in the fingerprint case, prove that adaptivity significantly improves the recognition system. So, ideally, we would characterize each class by the best wavelet packet tree that represents it. However, this is possible only if a suitable cost function can be found. Given that we have no

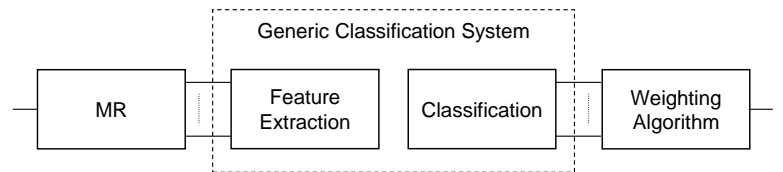


Figure 3. Our proposed adaptive MR classification system [13].

natural cost function available, we decided to mimic a wavelet-packet like system by adding a weighting procedure at the end of our system, allowing us to weigh the decisions of each subband in a fully grown tree. This way, a very low weight emulates a pruned branch in the tree. Thus, we propose a system with an MR decomposition block in front (see Fig. 3), followed by feature computation and classification in each of the subspaces, which are then combined through a weighting process, providing adaptivity.

Multiresolution Block. In our classification system, any MR transform can be used. In particular, amongst the MRBs, we used the DWT, DFT, Discrete Cosine Transform (DCT) and others, while amongst the MRFs, we used the Double-Density DWT (DD-DWT), Dual-Tree Complex Wavelet Transform (DT-CWT) and the Stationary Wavelet Transform (SWT), which is the most redundant transform. Note that here, we use all the subbands of the decomposition tree, not only the leaves. Thus, it might be abuse of language to call a transform a DWT. For example, for 2 levels, we have a total of $S = 21$ subbands (original image + 4 subbands at the first level + 16 subbands at the second level).

Feature Extraction and Classifier. We start with the feature sets used in [1]: Haralick texture features (set T_1 , 13 features), morphological (set M , 16 features) and Zernike moments (set Z , 49 features). Unlike in [1], we do not use wavelet/Gabor features because the MR advantage given by these will be achieved by our MR decomposition. Therefore, our total number of features is 78, as opposed to 174 in [1].

Instead of combining all features into a single feature vector, we allow each feature set its own feature vector per subband effectively bringing the number of subbands to $3 \cdot S = 63$ when using two levels of decomposition and all three feature sets. Note that although we have decreased the number of features significantly, we have also increased the number of classifiers, because we now have one classifier per subband. Evaluating this computational trade-off is a task for future work.

New Texture Feature Set T_3 . As we will show later on, the Haralick texture features seem to possess the most discriminative power, so we looked more closely into these. We changed the way that Haralick combines the initial four sets of features. We note that horizontal and vertical co-occurrence matrices are fundamentally different from the diagonal ones because adjacent neighboring pixels are spatially closer than diagonal neighboring pixels. Therefore, instead of averaging the features from all four sets, we create our first set of 13 features by averaging horizontal and vertical measures, and a second set of 13 features by averaging diagonal measures. Thus, we end up with a new feature set T_3 of 26 features [13].

Neural Networks. We decided to use a two-layer NN classifier. The first layer contains a node for each of the input features, each node using the Tan-Sigmoid transfer function. The second layer contains a node for each output and uses a linear transfer function (no hidden layers are used). In our design, when training, each output from the second layer corresponds to a class, and each training image will have an output of 1 for the class of which it is a member and a 0 for all other classes.

Weighting Procedure. Fig. 3 shows a graphical representation of a generic MR classification system, including the process of combining all of the subband decisions into one. We use weights for each subband to adjust the importance that a particular subband has on the overall decision made by the classification system. If the weights are chosen such that the no-decomposition weight is equal to 1, and all other weights are 0, we will achieve the same output vector as we would have without using the adaptive MR system. Therefore, we know that there exists a weight combination that will do at least as well as the generic classifier (when no MR is involved) in the training phase. Our goal is to decide how to find the weight vector that achieves the highest overall classification accuracy on a given dataset. We developed three versions of the weighting algorithm: open-form (OF), per-dataset closed-form (PD-CF) and per class closed-form (PC-CF). The PD-CF algorithm assigns one weight vector for the entire dataset, whereas the PC-CF assigns a weight vector for each class in the dataset. The latter goes back to our original idea of having a wavelet packet tree characterizing each class, only in this case, we do not necessarily obtain a tree.

The NN block outputs a series of decision vectors for each subband of each training image. Each decision vector $d_s^{(r)}$ contains C numbers (where C denotes number of classes) that correspond to the “local” decisions made by the subband s for a specific image r .

The classifier is evaluated using nested cross validations (five-fold cross validation in the NN block and ten-fold during the weighting process). One problem with this technique is that the initial ordering of the images determines which images are grouped together for training and testing in each fold of the cross validation. We solve this problem by running multiple trials, each with a random initial ordering of the images.

Open-Form Algorithm (OF). If using the OF algorithm, we initialize all the weights, and a global decision vector is computed using a weighted sum of the local decisions. An initial class label will be given to an image using this global decision vector. If that class label is correct, we go to the next image. If it is incorrect, we look at the local decisions of each subband and adjust the weights of each subband s as follows:

$$w_s^{iter} = \begin{cases} w_s^{iter-1} \cdot (1 + \epsilon) & \text{if subband } s \text{ is correct,} \\ w_s^{iter-1} \cdot (1 - \epsilon) & \text{otherwise,} \end{cases}$$

where $iter$ is the iteration number and ϵ is a small positive constant. This can be viewed as a reward/punishment method where the subbands taking the correct decisions will have their weights increased, and those taking wrong

decisions will have their weights decreased. We continue cycling through the images until there is no increase in classification accuracy on the training set for a given number of iterations.

Per-Dataset Closed-Form Algorithm (PD-CF). The CF solution does not use an iterative algorithm; rather, it finds the weight vector by solving a minimization problem in the least-square sense.

Assume we have R training images. For each training image $r = 1, \dots, R$, the vector $d_s^{(r)} = (d_{s,c}^{(r)})^T$ for classes $c = 1, \dots, C$, is the $C \times 1$ decision vector at the output of each subband classifier s , where $d_{s,c}^{(r)}$ indicates the confidence of subband s that the training image r belongs to class c . For each training image r , the weighting block takes as input the subband (local) decision vectors $d_s^{(r)}$ and combines them into a single output decision vector as follows:

$$\sum_{s=1}^S w_s d_s^{(r)}. \quad (12)$$

We can rewrite the above by, for each training image r , forming a matrix $D^{(r)}$ of size $C \times S$, where each element $D_{c,s}^{(r)}$ is the value at position c of the decision vector $d_s^{(r)}$ of subband classifier s . We can now compute $D^{(r)}w$, where $w = (w_1, \dots, w_S)^T$ is of size $S \times 1$. Thus, we want to find a weight vector w common to all training images $r = 1, \dots, R$.

A possible solution for w is the one that minimizes the error in the least-square sense:

$$w_{win} = \arg \min_w \sum_{r=1}^R \|d^{(r)} - D^{(r)}w\|^2, \quad (13)$$

where $d^{(r)}$ is the desired target decision vector of size $C \times 1$, with 1 in the position of the true class, and 0 elsewhere. We need to rewrite the above in a direct error-minimization form. We thus define a target output vector d of size $CR \times 1$, as a vector which concatenates all the target decision vectors $d^{(r)}$ as follows:

$$d = \left(\underbrace{d_1^{(1)}, d_2^{(1)}, \dots, d_C^{(1)}}_{\text{training image 1}}, \dots, \underbrace{d_1^{(R)}, \dots, d_C^{(R)}}_{\text{training image R}} \right)^T,$$

and a $CR \times S$ matrix D consisting of the all the decision matrices $D^{(r)}$ of all the training data stacked on top of each other. That is, $(D)_{cr,s} = D_{c,s}^{(r)}$ for $c = 1, \dots, C, r = 1, \dots, R$ and $s = 1, \dots, S$. We can now rewrite (13) as:

$$w_{win} = \arg \min_w \|d - Dw\|, \quad (14)$$

which possesses a CF solution and can be computed efficiently.

Then, for a testing image t , we compute its decision vector $\delta = (\delta_1, \delta_2, \dots, \delta_C)$ as follows:

$$\delta = \sum_{s=1}^S w_{win,s} d_s^{(t)},$$

where $d_s^{(t)}$ are the local decision vectors for t . The classification decision is then made as $c_{win} = \arg \max_c \delta_c$, that is, the winning class corresponds to the index of the highest coefficient in δ .

Per-Class Closed-Form Algorithm (PC-CF). To make the system truly adaptive, it is reasonable to assume that different classes require different weight vectors. Thus, we propose a system where, instead of a single weight vector w for the whole training dataset, each class c has its own weight vector w_c [16]. As opposed to (12), the entries in the output decision vector are now computed as:

$$D^{(r)}w_c, \quad c = 1, \dots, C. \quad (15)$$

Now, the weights can be grouped together to form an $S \times C$ matrix W so that each column represents a class-specific weight vector. Equation (15) can be rewritten as $\text{diag}(D^{(r)}W)$. Recall that $D^{(r)}$ is of size $C \times S$ and thus d is of size $C \times C$ (compare this to (12)). To learn these weights, we again use the training set and look for a solution that minimizes the squared error:

$$W_{win} = \arg \min_W \sum_{r=1}^R \|d^{(r)} - \text{diag}(D^{(r)}W^{(r)})\|^2. \quad (16)$$

To obtain an expression analogous to (14) and be able to apply standard methods, we have to define v as the vector concatenating all class-specific weight vectors:

$$v = (W_{1,1}, W_{1,2} \dots W_{1,C}, \dots, W_{S,1}, \dots, W_{S,C})^\top. \quad (17)$$

We now define D as the $CR \times CS$ block matrix $(D)_{rc,sc} = d_c^{(r)}$, where $d_c^{(r)}$ is the vector $(D_{c,1}^{(r)}, D_{c,2}^{(r)}, \dots, D_{c,S}^{(r)})$ and $r = 1, \dots, R$. We can now write a minimization problem equivalent to the one in (16), and which we can solve using standard techniques:

$$v_{win} = \arg \min_v \|d - Dv\|. \quad (18)$$

Decomposition Tree Pruning. Our long-term goal in developing an adaptive MR classification system was to find a WP-like decomposition, where each class would induce a different MR subtree. While the authors have done just that in [9], we needed a cost function which is specific to the dataset used. Our goal is thus to have a more generic system and to achieve a WP-like system but without the need for a cost function. We come close to this goal here, where we identify the set of discriminative subbands for each class (not necessarily a subtree).

Once the weight vectors are computed (using any version of the CF weighting algorithm), we use the values of the weights to regulate the MR decomposition. In particular, subbands with low weight can be pruned as long as the remaining subbands are still sufficient to classify the image correctly. This way, the pruned subbands and their associated features need not be computed, resulting in computational savings (although not increased accuracy). We propose to keep the high-weight subbands, so that at least a certain ratio, defined as the fraction of the sum of kept weights over the sum of all the weights, of subbands are kept.

3. CLASSIFICATION PROBLEMS IN BIOIMAGING AND BIOMETRICS

While we have developed the current algorithm by learning from each application as we went along, we decided to first present all algorithmic accomplishments and then discuss results in various application domains. We do that now and use different instantiations of the MR classification algorithm depending on the dataset at hand.

Determination of Protein Subcellular Location Patterns. To evaluate our MR approach, we use the 2D HeLa set depicting PSL described previously [1]. The proteins in the dataset were labeled using immunofluorescence, and thus, we know the ground truth, that is, which protein was labeled in each cell and subsequently imaged. The details of our results in this area can be found in [13].

The challenge in this dataset is that images from the same class may look different while those from different classes may look very similar (see Fig. 4(a)). This dataset is publicly available [17] and contains 50 single-cell images of size 512×512 , in each of $C = 10$ classes. The 10 classes of subcellular location patterns were obtained by labeling an endoplasmic reticulum protein, two Golgi proteins (giantin and gpp130), a lysosomal protein, a mitochondrial protein, a nucleolar protein, two cytoskeletal proteins (actin and tubulin), an endosomal protein, and DNA. The best previously described overall classification accuracy on this dataset is 91.5% [1]. We achieve the best classification accuracy of 95.40% using MRFs (see Fig. 5(a)) [13].

Detection of Developmental Stages in *Drosophila* Embryos. The details of our results in this area can be found in [14]. The dataset consists of 60 time-lapse, fluorescence microscopy z-stacks (3D volumes in time) of developmental stages of *Drosophila* embryos (see Fig. 4(b)). The stacks are acquired roughly every 10 minutes. The number of slices per stack varies; it is 5 slices for normal sets and 7 slices for delayed/abnormal. The number of time points is typically 15 for normal/abnormal and around 30 for delayed. All the slices have been tagged by

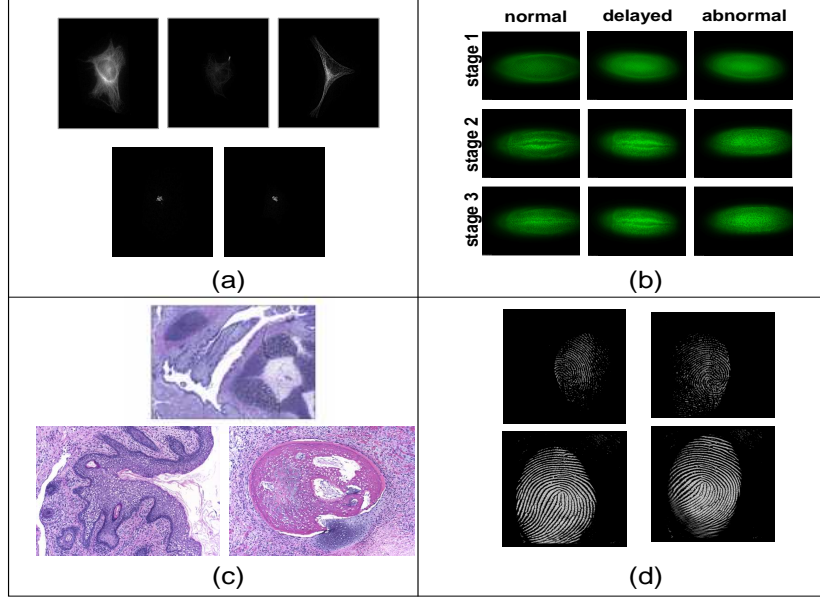


Figure 4. (a) Top: Intra-class variation. The three images show the spatial distribution of Tubulin within a cell; Bottom: Interclass similarity. The first image shows the spatial distribution of Giantin and the second image shows the spatial distribution of Gpp130. Both are Golgi-proteins. (Images courtesy of [17]). (b) Representative examples of each stage. Top: Stage 1, no ventral furrow, for normal ($t=30\text{min}$), delayed ($t=60\text{min}$) and abnormal ($t=20\text{min}$) embryos. Middle: Stage 2, ventral furrow opening, for normal ($t=60\text{min}$), delayed ($t=110\text{min}$) and abnormal ($t=72\text{min}$) embryos. Bottom: Stage 3, ventral furrow closed, for normal ($t=75\text{min}$), delayed ($t=140\text{min}$) and abnormal ($t=82\text{min}$) embryos [2]. (c) Examples of histological images depicting stem cells differentiating into bronchus (top), skin (bottom left) and bone (bottom right) [15]. (d) Example fingerprint images from a difficult class (top) and an easy class (bottom) [18].

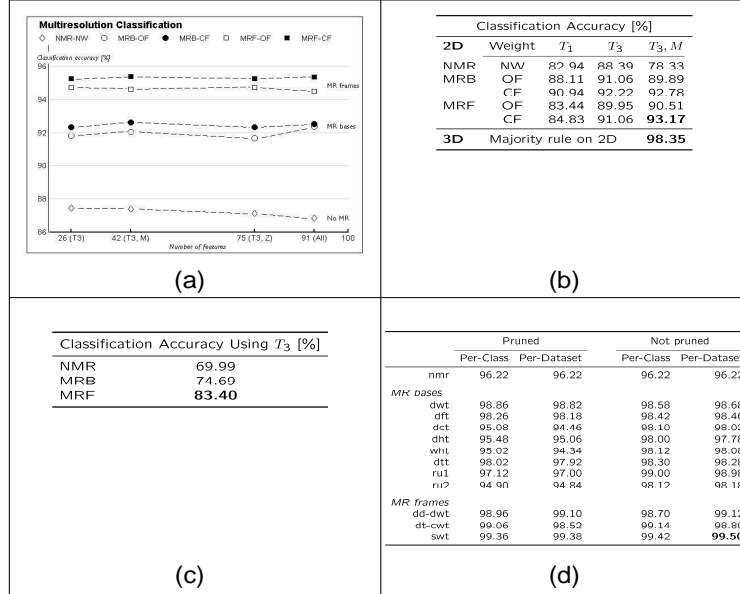


Figure 5. Classification results for all four datasets. We indicate in bold the highest accuracy achieved by each transform. (a) Pictorial representation of classification accuracy for 2D HeLa images depicting PSL patterns [13]. (b) Classification accuracy for 2D slices of Drosophila embryos. We use these in majority voting classification for 3D stacks yielding the accuracy of 98.35% [14]. (c) Classification accuracy for stem cell teratomas [15]. (d) Classification accuracies for fingerprint images obtained with different MR transforms using two weighting algorithms and a pruning procedure [16].

a human expert so we have reliable ground truth. As depicted in Fig. 5(b), the highest classification accuracy achieved by our system is 93.35% (see details in [14]).

Classification of Histological Stem-Cell Teratomas. Here, we first populate the dataset with a fair amount of single-class images. We start with H&E images (at 10x magnification, see Fig.4(c)) that depict multiple tissues (classes) contained in the teratomas. These are hand segmented by an expert to separate the classes. Then, the segmentation masks are used to generate single-class images. Because the set of multi-class images available to us is small (only 23 images), we decided to take advantage of their large size (1600×1200), and use a window to extract single-class images of size 200×200 . We thus obtain 45 images per class. We use six classes for this experiment: mesenchyme (embryonic connective tissue), skin, myenteric plexus, bone, necrotic (dying or dead tissue), and striated muscle. The best classification accuracy of 83.40% is achieved by MRFs (see Fig. 5(c)) [15].

Fingerprint Recognition. To test our system we used images from a subset of the NIST 24 fingerprint database [18]. The dataset contains 10 classes with $50 \times 512 \times 512$ images each (45 images are used to train the system). The images were acquired while individuals were rolling their thumbs, inducing different plastic distortions making the dataset realistic and challenging (see Fig. 4(d)). As shown in Fig. 5, we achieve the highest accuracy of 99.50% by using the SWT along with the PD-CF weighting algorithm [16].

3.1. Results and Discussion

All the results are shown in Fig. 5. By observing the results for all applications, we can draw the following conclusions (note that NMR denotes the version of the algorithm where no MR transform is used):

- In all cases, MR significantly outperforms NMR, thus demonstrating that classifying in MR subspaces indeed improves classification accuracy.
- The redundant transforms (MR frames) do better than nonredundant ones (MR bases). In all cases, the SWT achieves the best classification accuracy.
- While a slightly higher classification accuracy is obtained by using all three feature sets as well as both T and M , the larger number of features and additional complexity of using M and Z features do not justify the slight improvement in accuracy. This “flat” trend (see Fig. 5(a)) is good news as we can use a significantly reduced feature set and still obtain a fairly high classification accuracy.
- The closed-form version of the weighting process gives slightly better results than the open-form.
- As expected, when used, while pruning does not improve the accuracy of the system, it does make the system more efficient.
- In general, the class-adaptive method seems to do better than the dataset-adaptive one.

4. DESIGN OF NEW FRAME FAMILIES

Examining the results we obtained with our MR classification system, we found the trends to be similar: MR significantly outperforms NMR and the best results are invariably obtained by frames. Whether it is classifying biological or biometric images, frames and in particular the SWT always outperformed any other transform. Taking into consideration the computational cost, it is important to have a system that is efficient in addition to being accurate. The SWT is the most accurate here but also the most redundant. Therefore, to allow for a trade-off between accuracy and cost, we would like to create new redundant transforms that are less redundant but still afford very good accuracies when it comes to classification.

A known issue with MRBs is that they are not translation invariant (rather, they are periodically translation invariant). This is due to downsampling being used and can create problems as translated versions of data can lead to different features in MR subspaces. As for the fingerprint dataset, we conducted an experiment on the PSL images to test the sensitivity of our classification system to translations. We used T_3 features, trained the algorithm on the original data and tested on translated PSL images. As expected, the classification accuracy dropped by 0.22%. Both experiments for fingerprints and PSL strongly indicate the use of MR techniques which are translation invariant (or almost translation invariant).

These considerations lead us to believe that properties provided by frames, on top of the MR ones, are crucial requirements in some applications. Motivated by the need of having frame families dedicated to a spectrum of applications not considered before, we seek to design new classes of frames. Details can be found in [19].

	finite-dimensional (block transforms)		infinite-dimensional (overlapped transforms)
ONBs	DFT	→	LOT
	↓		↓
TFs	HTF	→	LTFT

The question now is: How do we go about constructing new families and what do we look for? Most of the known frame families (though not all) are block ones (finite dimensional) leading to blocking effects. We want to have efficient implementations as well as be able to flexibly decide on the requisite amount of redundancy. These requirements made us think of LOTs: As mentioned in Section 1.3, on top of being computationally efficient, the LOTs have the advantage of processing blocks of overlapping data and hence eliminate blocking artifacts. So the question is: Could we construct a similar transform with frames? Our idea is to seed LOTs to obtain a new class of frames we name *Lapped Tight Frame Transforms (LTFTs)*. That is, we want to find filter-bank frames seeded from the LOTs in the hope they will inherit all the good properties LOTs possess. Obtained by seeding, the LTFTs could thus be seen both as the frame counterpart of LOT bases as well as the infinite-dimensional, filter-bank counterpart of the most famous frame family—Harmonic Tight Frames (HTFs, seeded from the DFT). These relationships are illustrated in the table.

An effort to derive overlapped frame transforms (not with seeding) can be found in [20]. These are not obtained by seeding (they start from a frame) and while are in spirit similar to what we are proposing, they lead to a completely different family. The same authors have already proposed a 2D nonseparable frame family.

4.1. Lapped Tight Frame Transforms.

We propose a host of new frame families we denote as *Lapped Tight Frame Transforms (LTFTs)* (details about this part of our work are given in [19]):

DEFINITION 4.1. *A lapped tight frame is a frame transform (LTFT) seeded from an LOT.*

Let us start with $\Psi_p(z)$ being the $m \times m$ polyphase matrix associated with the LOT of size m . Then (4) holds and we seed the LOT to get:

$$\Phi_p^*(z) = \Phi_0^* + z^{-1}\Phi_1^* = \Psi_p[\mathbb{J}]. \quad (19)$$

The matrices Φ_r^* are now rectangular of size $n \times m$. For $r = 0, 1$, we have $(\Phi_r)_{i,j} = \psi_{i,mr+j}^*$, with $i = 0, \dots, n-1$ and $j = 0, \dots, m-1$. By Naimark Theorem, we know that this family is a TF, which implies that $\Phi_p(z)\Phi_p^*(z) = cI$ (c is a constant). Note that as opposed to the LOT case, the matrix products do not commute anymore.

Princen-Johnson-Bradley LTFTs. All of the above is general and can be applied to any type of LOT. Let us now see through an example what happens when the obtained LTFT has been seeded by the PJB-LOT in (7). We will call these *Princen-Johnson-Bradley LTFTs (PJB-LTFTs)*.

Seeding. From the Naimark theorem, we know that any general seeding will result in a TF. However, if we want more than tightness (equal-norm, for instance), in general, there is no guarantee that we will obtain such a property. One needs to carefully choose the set of retained columns to preserve those properties. Indeed, seeding the DFT leads to HTFs only if the set of eliminated columns is contiguous. Choosing non contiguous columns leads to tight but not equal-norm frames. In the PJB case, it turns out that we can have a general seeding and any subset of columns (not necessarily contiguous) can be retained to obtain an ENTf. We summarize this result in the following (the proof is technical and not included here):

LEMMA 4.2. *The PJB-LTFTs obtained by any seeding of the PJB-LOTs are ENTfs. That is, $\Phi_p^*(z) = \Psi_p[\mathbb{J}]$ for any subset \mathbb{J} of columns is an ENTf, with norm $\sqrt{n/m}$.*

For PJB-LOTs we can compute $\text{diag}(\Phi_0^*\Phi_0 + \Phi_1^*\Phi_1)_i = \|\varphi_i\|^2$. From Lemma 4.2, we know that without loss of generality we can assume that $\mathbb{J} = 0, \dots, n-1$. Note that

$$\|\varphi_i\|^2 = \sum_{j=0}^{n-1} \psi_{j,i}^{*2} + \psi_{j,i+m}^{*2}.$$

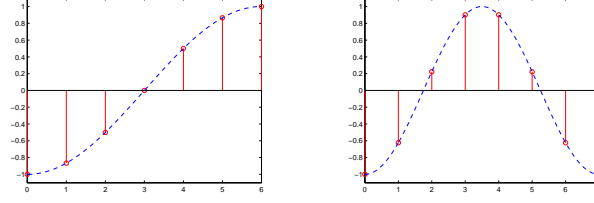


Figure 6. Window solution to (24)-(25) for $n = 7, 8$ (left to right).

In fact, we can find the equal norm as $\|\varphi_i\|^2 = n/m$, for $i = 0, \dots, m-1$. That is, the LTFT obtained is equal-norm.

Window Design. If we start with the PJB-LOT with a window, and seed $W\Psi$, the TF obtained would lose its equal-norm property since $\|\varphi_i\|^2 = (n/m)w_i^2$. To preserve equal norm, we have to modulate directly the LTFT after seeding the LOT. In the PJB-LOTs, the window chosen was symmetric, that is, $w_i = w_{2m-1-i}$. We lift this restriction initially and assume a general window represented by a matrix D , a $2n \times 2n$ diagonal matrix. We can write $D = (D_0 \ D_1)$ and D_r is a $n \times n$ diagonal matrix. Unlike for the LOTs, the matrix product $\Phi_0\Phi_0^*$ has no particular structure, in fact,

$$(\Phi_0\Phi_0^*)_{i,j} = a_{i,j} = \frac{1}{2m} \frac{\sin(\frac{\pi(i+j+1)}{2})}{\sin(\frac{\pi(i+j+1)}{2m})} + \frac{1}{2m} \frac{\sin(\frac{\pi(i-j)}{2})}{\sin(\frac{\pi(i-j)}{2m})}.$$

Substituting this into (5), we obtain the following:

$$a_{j,j}d_j^2 + (1 - a_{j,j})d_{n+j}^2 = 1, \quad (20)$$

$$d_j d_s = d_{n+j} d_{n+s}, \quad s = 0, \dots, n-1, s \neq j. \quad (21)$$

The set of solutions to (20)-(21) is infinite. Of course, the constant window with $d_j = 1$, for $j = 0, \dots, 2n-1$ is also a solution to the above. If the window is symmetric, then (5) becomes:

$$D_0\Phi_0\Phi_0^*D_0 + JD_0J\Phi_1\Phi_1^*JD_0J = I \quad (22)$$

$$\text{with } \Phi_0\Phi_0^* + \Phi_1\Phi_1^* = I. \quad (23)$$

Using (22), we derive the following conditions on D :

$$a_{j,j}d_j^2 + (1 - a_{j,j})d_{n-j-1}^2 = 1, \quad (24)$$

$$d_j d_s = d_{n-j-1} d_{n-s-1}, \quad s = 0, \dots, n-1, s \neq j. \quad (25)$$

Fixing $d_0 = -1$, we have $d_{n-1} = \pm 1$ and $d_s = -d_{n-1}d_{n-s-1}$ for $s = 1, \dots, n-2$. Note that the same conditions hold for an anti-symmetric window, that is, the half-windows can only be symmetric or antisymmetric. For a symmetric window, a possible solution, depicted in Fig. 6, is given by

$$d_j = \begin{cases} \cos(\frac{j\pi}{n-1} + \pi) & \text{if } n \text{ is even,} \\ \cos(\frac{2j\pi}{n-1} + \pi) & \text{if } n \text{ is odd,} \end{cases} \quad j = 0, \dots, n-1.$$

One of our first tasks in future work will be to use these symmetric windows in the PJB-LTFTs and study their effect. We also need to investigate window design techniques to obtain optimized windows that will modulate the PJB-LTFT and hence lead to better localization in the frequency band of the frame vectors.

REFERENCES

1. K. Huang and R. Murphy, "Boosting accuracy of automated classification of fluorescence microscope images for location proteomics," *BMC Bioinformatics* **5**(78), 2004.
2. Z. Kam, J. Minden, D. Agard, J. Sedat, and M. Leptin, "Drosophila gastrulation: Analysis of cell shape changes in living embryos by three-dimensional fluorescence microscopy," *Development* **112**, pp. 365–370, 1991.
3. M. Adams, S. Celniker, R. Holt, C. Evans, J. Gocayne, P. Amanatides, S. Scherer, P. Li, R. Hoskins, R. Galle, and et al., "The genome sequence of *Drosophila melanogaster*," *Science* **287**, pp. 2185–2195, 2000.
4. R. Haralick, "Statistical and structural approaches to texture," *Proc. IEEE* **67**, pp. 786–804, 1979.
5. M. Vetterli and J. Kovačević, *Wavelets and Subband Coding*, Signal Processing, Prentice Hall, Englewood Cliffs, NJ, 1995.
6. J. Kovačević and A. Chebira, "Life beyond bases: The advent of frames (Part I)," *IEEE Signal Proc. Mag.* **24**, pp. 86–104, Jul. 2007.
7. H. Malvar, *Signal Processing with Lapped Transforms*, Artech House, Norwood, MA, 1992.
8. J. Princen, A. Johnson, and A. Bradley, "Subband transform coding using filter bank designs based on time domain aliasing cancellation," in *Proc. IEEE Int. Conf. Acoust., Speech and Signal Proc.*, pp. 2161–2164, (Dallas, TX), Apr. 1987.
9. P. H. Yeomans, J. Thornton, J. Kovačević, and B. Kumar, "Wavelet packet correlation methods in biometrics," *Appl. Opt., sp. iss. Biometric Recognition Systems* **44**, pp. 637–646, Feb. 2005.
10. J. Kovačević and A. Chebira, "Life beyond bases: The advent of frames (Part II)," *IEEE Signal Proc. Mag.* **24**, Sep. 2007.
11. Z. Cvetković and M. Vetterli, "Oversampled filter banks," *IEEE Trans. Signal Proc.* **46**, pp. 1245–1255, May 1998.
12. M. Püschel and J. Kovačević, "Real, tight frames with maximal robustness to erasures," in *Proc. Data Compr. Conf.*, pp. 63–72, (Snowbird, Utah), Mar. 2005.
13. A. Chebira, Y. Barbotin, C. Jackson, T. Merryman, G. Srinivasa, R. Murphy, and J. Kovačević, "A multiresolution approach to automated classification of protein subcellular location images," *BMC Bioinformatics* **8**(210), 2007.
14. R. Kellogg, A. Chebira, A. Goyal, P. Cuadra, S. Zappe, J. Minden, and J. Kovačević, "Towards an image analysis toolbox for high-throughput *Drosophila* embryo RNAi screens," in *Proc. IEEE Int. Symp. Biomed. Imaging*, pp. 288–291, (Arlington, VA), Apr. 2007.
15. J. Ozolek, C. Castro, W. Jenkinson, A. Chebira, J. Kovačević, C. Navara, M. Sukhwani, K. Orwig, A. Ben-Yehudah, and G. Schatten, "Semiquantitative and multiresolution-based histological analysis of germ layer components in teratomas derived from human, non-human primate and mouse embryonic stem cells," (Cairns Australia), Jun. 2007.
16. A. Chebira, L. Coelho, A. Sandryhalia, S. Lin, G. Jenkinson, J. MacSleyne, C. Hoffman, P. Cuadra, C. Jackson, M. Püschel, and J. Kovačević, "An adaptive multiresolution approach to fingerprint recognition," in *Proc. IEEE Int. Conf. Image Proc.*, (San Antonio, TX), Sep. 2007. To appear.
17. The Murphy Lab at Carnegie Mellon University: <http://murphylab.web.cmu.edu>.
18. C. Watson, "NIST Special Database 24: Live-Scan Digital Video Fingerprint Database." National Institutes of Health, Gaithersburg, Md., 1998. <http://www.nist.gov/srd/nistsd24.htm>.
19. A. Chebira and J. Kovačević, "Lapped tight frame transforms," in *Proc. IEEE Int. Conf. Acoust., Speech and Signal Proc.*, pp. 857–860, (Honolulu, HI), Apr. 2007.
20. J. Gauthier, L. Duval, and J. Pesquet, "Low redundancy oversampled lapped transforms and application to 3D seismic data filtering," in *Proc. IEEE Int. Conf. Acoust., Speech and Signal Proc.*, **II**, pp. 821–824, (Toulouse, France), May 2006.